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DEVELOPING TECHNOLOGY FOR LONG-TERM HOLDING OF MUSSELS IN CAPTIVITY

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Quarantine Experiments

In 1996 and 1997, mussels were quarantined under static conditions in 1,136 liter stock tanks. These tanks were equipped with aerators (1/20 hp). After quarantine, all water in the tanks was treated with calcium hypochlorite for ≥ 5 hours to kill all organisms, including any zebra mussels. Following this treatment all tanks were hand scrubbed and rinsed several times with tap water prior to use for another quarantine. Thus, this treatment likely eliminated most bacteria. During each quarantine, total ammonia began to increase immediately following introduction of mussels. Typically, maximum concentrations of 0.5 to 3.2 mg/L were reached in 3 to 17 days. Ammonia levels then declined gradually and remained low for the remainder of the quarantine. We attributed this decline in ammonia to the build-up of denitrifying bacteria on the tank surfaces as well as on the mussel shells. Moreover, maximum ammonia concentrations seemed to be negatively related to mussel survival during quarantine and survival during the following 12 months at the permanent holding facility.

In 1998, we designed and constructed a two-tier recirculating system for quarantining mussels. The system includes two of the stock tanks used in previous years, a sump, and a biofilter. Water is pumped from the lower tank up to the sump. An overflow pipe in the sump drains into the biofilter. The water is then gravity-fed into the upper tank, and then down into the lower tank. About two weeks before quarantining mussels, we innoculated the biofilter with filter material from another recirculating system used for

fish. At the same time, we introduced about 50 fish into the quarantine tanks to maintain the bacterial culture.

In September 1998, we collected mussels from the Diamond Island area of the Tennessee River, and hand-scrubbed them to remove any zebra mussels. Prior to quarantine, all fish were removed from the recirculating system, and then mussels were introduced at a density of 1 mussel/3.4 liters of water (for the entire system). Ammonia levels remained below 0.25 mg/L, and generally were undetectable. Survival of all species was > 83% at the end of the quarantine (Table 1). Following quarantine, these mussels were translocated to the French Broad River.

Because the recirculating system was so successful, we built a second system. At the same time, we began culturing algae (*Bracteococus grandis*) to feed mussels during quarantine. In October 1998, we again collected mussels near Diamond Island. We followed the same protocol used for the September quarantine with the following exceptions. Mussels were quarantined in one recirculating system at the same density (1 mussel/3.4 L) used in September; the density was doubled (1 mussel/1.7 L) in the second recirculating system. Throughout the quarantine period mussels in both recirculating systems were fed live algae at a rate of 1.4 mg (dry weight) per mussel every 2-3 days. Overall mussel survival exceeded 95% in both recirculating systems (Table 1). Following quarantine, these mussels were moved to our long-term holding facility at Center Hill Reservoir. Survival of these mussels will be monitored for the next year.

Physiological Condition

Biochemical analysis of tissue samples of Fusconaia ebena and Amblema plicata were conducted to assess physiological condition of these species in Kentucky Lake at the beginning and end of each quarantine period in 1997. Similar analyses were performed on samples of these species following quarantine. Four replicates of a composite sample of 10 individuals of each species were analyzed for percent lipid composition. In most cases, 11/1 there was no apparent affect of quarantine on percent lipid composition following quarantine. Similarly, protein composition varied little among treatments; however, there was a highly significant loss in biomass during most quarantines and densities. In general, mean dry tissue weight adjusted ghall and for shell length for quarantined mussels was 11 to 32% less than for individuals of F. ebena not quarantined. Although dry tissue weights of A. plicata also decreased 5 to 40%, most of these differences were not significant. These data are being subjected to further statistical analysis.

Captive Populations

During 1998, we consolidated our captive mussel populations into three facilities: Minor E. Clark Hatchery, KY; the Normandy Hatchery, TN; and Center Hill Reservoir. Mussels previously held at the Laurel Hill Wildlife Management Area, Elkhorn Station, and the Birdsong Marina (American Pearl Farm) were either moved to one of these facilities or were returned to their natal stream. Additionally, some captive mussels were translocated to the

French Broad River. As of December 31, 1998 we have 8,057 mussels of 49 species in captivity (Table 2).

In 1996, we collected 100 mussels of 6 species from Elkhorn Creek near the Frankfort Hatchery. These mussels were transported to Elkhorn Station and placed into pocket nets suspended in an old raceway/pond. Initial (30-day) survival was 100% (Table 3). Survival after 2 years was high for most species. Moreover, some individuals of all species of *Lampsilis* became gravid while being held in this facility. In 1998, all mussels were returned to Elkhorn Creek. We believe that this facility is suitable for maintaining refuge populations of unionids should the need arise.

In 1994, we collected 272 mussels of 7 species from the Licking River and placed them into a raceway at the Minor E. Clark Hatchery. Additional mussels were introduced in 1995, 1996, and 1998. Most of the species became gravid in the year following their introduction, as well as in subsequent years. As part of another study on mussel propagation, juvenile *Lampsilis cardium*, produced in one of the raceways, reached a mean length of 21.5 mm after one growing season and 54.5 mm after the second growing season, and about 75 mm after the third season. Because the parental stock (4 gravid females) came from Buck Creek, KY, we have stocked 400 of these cultured *Lampsilis cardium* (1½ to 2 yr old) into the creek. The remaining individuals (>400) are in captivity and will be stocked in different locations to enhance or reestablish mussel populations. Thus,

this facility is well-suited not only to maintain refuge populations of mussels, but also for the propagation and culture of mussels.

Survival of mussels at the Normandy Hatchery has been excellent. In addition to common species, we are holding (under the necessary Federal Endangered Species Permits) 87 mussels of 7 endangered species (Table 1). This facility has a lot of potential for the propagation and culture of endangered species. In 1995, we propagated *Lampsilis fasciola* in the raceway and have continued culturing this cohort. In September 1998, we discovered that these juveniles had spawned and became gravid. To the best of our knowledge, this represents the only instance of culturing mussels through their entire life cycle since Howard (1916).

Reference

Howard, A.D. 1916. A second generation of artificially reared fresh-water mussels. Transactions of the American Fisheries Society 46:93-100.

Table 1. Survival of mussels during a 30-day quarantine in recirculating systems in 1998.

	Quarantine Density (liters of water per mussel)	Species and Percent Survival			
Month Collected		Cyclonaias tuberculata	Quadrula metanevra	Quadrula pustulosa	
September	3.4/1	98.9	100	83.9	
October	3.4/1	99.5	100	92.2	
October	1.7/1	99.5	100	95.1	

Table 2. Species and numbers of mussels in captivity as of December 31, 1998.

	Facility		
Species	Normandy Hatchery	Minor Clark Hatchery	Center Hill Reservoir
Actinonaias ligamentina		142	
Actinonaias pectorosa	23	40° 400	
Amblema plicata		440 AM 444,	479
Arcidens confragosus			9
Cyclonaias tuberculata		Rph 1409-2009	905
Cyprogenia stegaria	1		
Elliptio crassidens	***	one was look	133
Elliptio dilatata		240	··-·
Ellipsaria lineolata			237
Epioblasma capsaeformis	20	MA AND THE	*** **** ****
Fusconaia ebena		#\$4 400 MGD	2255
Fusconaia flava		MA, ANA. 1004	126
Fusconaia subrotunda		3	*****
Lampsilis abrupta	37	main comp subpe	
Lampsilis cardium		478	AND SIDE WAS
Lampsilis fasciola	106	- 44 m	Note that with
Lampsilis ovata	12	**** ****	Article states
Lasmigona costata		5	
Lemiox rimosus	10		
Leptodea fragilis	and also take		1
Ligumia recta	62	ALL ART WAS	6
Medionidus conradicus	23		
Megalonaias nervosa	Marin 1969-4825		159
Obliquaria reflexa	and the Gar		34
Obovaria olivaria	ang pain angs		32
Plethobasus cicatricosus	4		446 distribu
Plethobasus cooperianus	14		AMP NOTE OFF
Pleurobema cordatum	13	158	71
Pleurobema rubrum		8	err mande
Pleurobema sintoxia		27	
Potamilus alatus		MAN AND 2,300	10
Pyganodon grandis	with final data	1	MAR WAR 400
Ptychobranchus fasciolaris	And 440 994	106	1
Ptychobranchus subtentum	14	Ann hab, gap,	into sale sub-
Quadrula apiculata	tor majo idan		1

Table 2 (continued).

	Facility			
Species	Normandy Hatchery	Minor Clark Hatchery	Center Hill Reservoir	
Quadrula cylindrica	8	ma 444 ma		
Quadrula metanevra	494 100 100	- np. sp.	721	
Quadrula nodulata	est also take	nun Any yan	59	
Quadrula pustulosa	dead white work	THE SAN SILV	1197	
Quadrula quadrula	and two and		16	
Tritogonia verrucosa		50	3	
Truncilla truncata	er to as		2	
Utterbackia imbecillis	while from your	1		
Villosa iris	30			
Villosa taeniata		1		
Villosa trabalis	1	man anga anga	**************************************	
Villosa vanuxemensis	3	an việt giệc	And the New	
Total	381	1219	6457	

Table 3. Numbers and percent survival of mussels collected July 9, 1996 from Elkhorn Creek and held at Elkhorn Station, KY.

		Percent Survival		
Species	Number Held	30-day	1-year	2-year
Amblema plicata	28	100	96	79
Elliptio dilatata	44	100	98	84
Lampsilis cardium	5	100	100	60
Lampsilis fasciola	2	100	100	100
Lampsilis siliquoidea	15	100	87	80
Lasmigona costata	6	100	83	50